

JC-1 - Mitochondrial Membrane Potential Assay Kit ab113850

★★★★☆ 5 Abreviews 83 References 画像数 6

医薬用外劇物

製品の概要

| | |
|----------|---|
| 製品名 | JC-1 - Mitochondrial Membrane Potential Assay Kit |
| 検出方法 | Fluorescent |
| サンプルの種類 | Adherent cells, Suspension cells |
| 全工程の試験時間 | 1h 00m |
| 製品の概要 | <p>JC1- Mitochondrial Membrane Potential Assay Kit ab113850 contains tetraethylbenzimidazolylcarbocyanine iodide (JC-1), a cationic dye that accumulates in energized mitochondria.</p> <p>At low concentrations (due to low mitochondrial membrane potential), JC-1 is predominantly a monomer that yields green fluorescence with emission of 530 ± 15 nm.</p> <p>At high concentrations (due to high mitochondrial membrane potential), the dye aggregates yielding a red to orange colored emission (590 ± 17.5 nm).</p> <p>Therefore a decrease in the aggregate fluorescent count is indicative of depolarization whereas an increase is indicative of hyperpolarization.</p> <p>The JC-1 staining protocol is very simple:</p> <ul style="list-style-type: none"> - wash cells in dilution buffer or PBS - add JC solution - incubate for 30 min at 37°C for suspension cells, or 10 min for adherent cells - wash cells with dilution buffer - treat cells as desired for experimental plan - analyze on a fluorescent microplate reader <p>The aggregate dye can be excited at 535 nm, the monomer and aggregate together at 475 nm.</p> |
| 特記事項 | <p>Review our cell health assays guide to learn more about our other cell viability, cytotoxicity and cell proliferation assay kits.</p> <p>Review the metabolism assay guide to learn about assays for metabolites, metabolic enzymes, mitochondrial function, and oxidative stress, and also about how to assay metabolic function in live cells using your plate reader.</p> |

製品の特性

保存方法

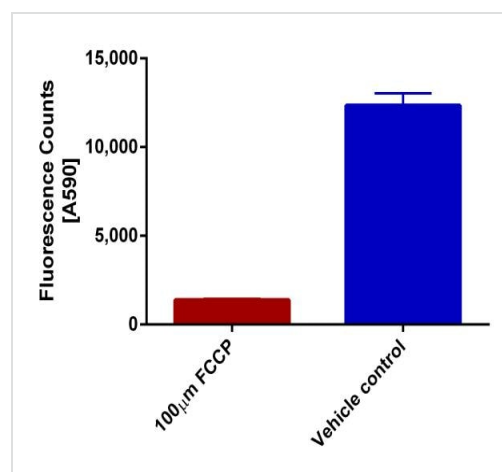
Store at -20°C. Please refer to protocols.

| 内容 | 100 tests |
|--------------------------------|-----------------|
| 50mM FCCP (in DMSO) | 1 x 10 μ l |
| Dilution Buffer (10X, sterile) | 1 x 10ml |
| DMSO (cell culture tested) | 1 x 1ml |
| JC-1 (lyophilized) | 1 x 500 μ g |

関連性

Mitochondrial Membrane Potential is an important parameter of mitochondrial function used as an indicator of cell death. The collapse of the mitochondrial Membrane potential coincides with the opening of the mitochondrial permeability transition pores, leading to the release of cytochrome c into the cytosol, which in turn triggers other downstream events in the apoptotic cascade.

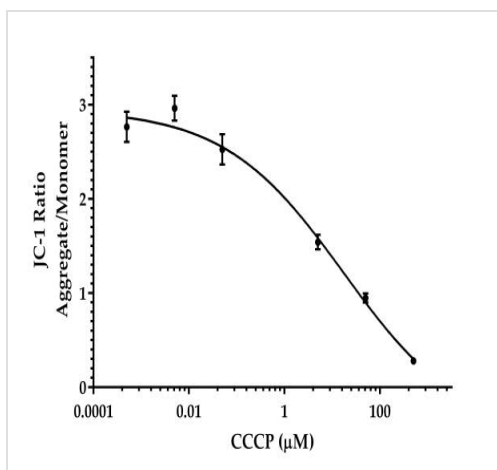
画像



JC-1 assay result in HL60 cells treated with FCCP

JC1 - Mitochondrial Membrane Potential Assay Kit (ab113850).

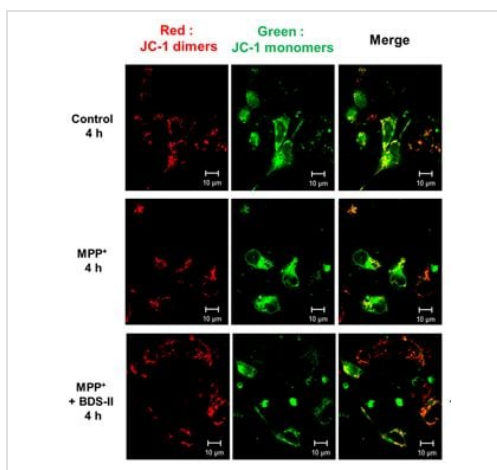
HL60 cells were seeded and labeled according to section 11.1 of the protocol. Cells were then treated for 4 hours with 100 μ M FCCP or vehicle/diluent control (DMSO). Mean and standard deviation is plotted for 3 replicates from each condition.



JC-1 assay result in HepG2 cells treated with CCCP.

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HepG2 cells were seeded and labeled according to section 11.2 of the protocol. Cells were then treated for 4 hours with a titration series of CCCP (carbonyl cyanide 3-chlorophenylhydrazone) and both monomer and aggregate forms were read on a Perkin Elmer-Wallac 1420 Victor 2 Multilabel plate reader. Mean and standard deviation of aggregate/monomer ratios is plotted for 12 replicates for each concentration. IC₅₀ of CCCP in HepG2 cells was calculated at 8.7 μM.



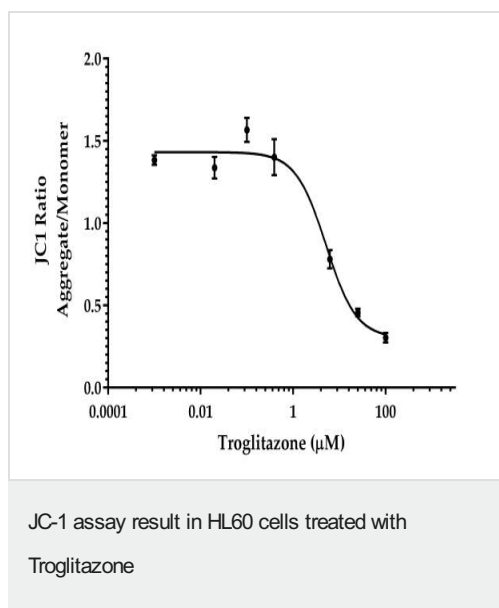
Using JC1 dye to examine mitochondrial membrane potential depolarization in SH-SY5Y cells

Image courtesy of Son MS et al. Sci Rep. 2017; 7: 2075. doi: 10.1038/s41598-017-02129-w. Reproduced under the Creative Commons License <http://creativecommons.org/licenses/by/4.0/>.

Son MS et al. (2017) used JC1 Mitochondrial Membrane Potential Assay Kit ab113850 to stain:

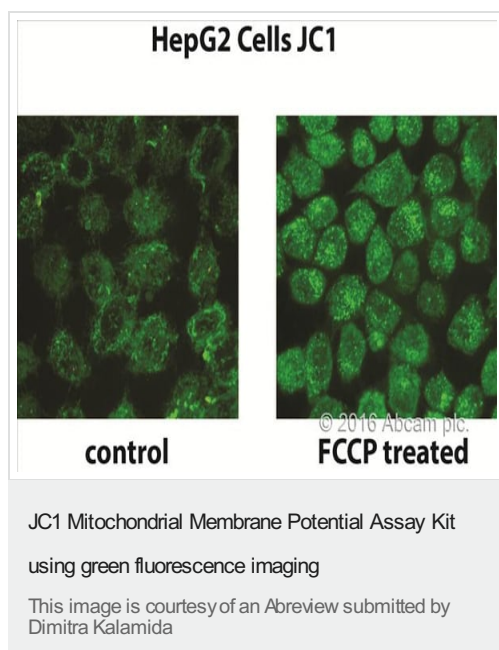
- untreated SH-SY5Y cells (control),
- SH-SY5Y cells treated with 1mM MPP⁺ (MPP⁺) and,
- SH-SY5Y cells pretreated with BDS-II followed by 1mM MPP⁺ treatment (MPP⁺ +BDSII).

Normal mitochondrial membrane potential is shown in red with JC-1 dimers and depolarized membrane potential is shown in green in JC-1 monomers.



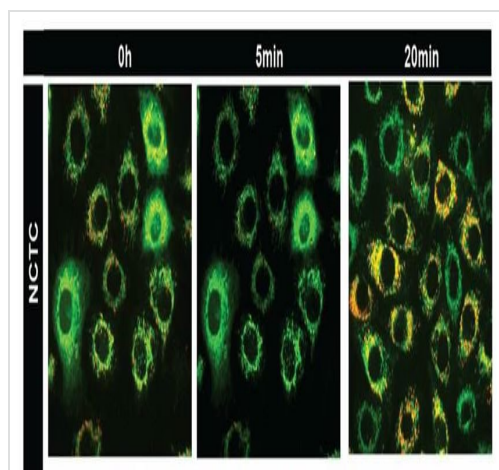
JC1 - Mitochondrial Membrane Potential Assay Kit (ab113850).

HL60 cells were seeded and labeled according to section 11.1 of the protocol. Cells were then treated for 4 hours with a titration series of the thiazolidinedione Troglitazone and both monomer and aggregate forms were read on a Perkin Elmer-Wallac 1420 Victor 2 Multilabel plate reader. Mean and standard deviation of aggregate/monomer ratios is plotted for 3 replicates for each concentration. IC₅₀ of Troglitazone in HL60 cells was calculated at 1.2 μM.



The JC1- Mitochondrial membrane potential assay kit has been tested using HepG2 cells, control cells and FCCP-treated cells (100μM for 4h) have been used as a positive control. The company's instructions were followed for JC1 mitochondrial membrane potential assay. Imaging was performed on a customized Andor Revolution Spinning Disk Confocal System built around a stand (IX81 Olympus) with a 60x lens and a digital camera (Andor Ixon+885) (CIBIT Facility, MBG-DUTH). Image acquisition was performed in Andor IQ 2 software. Optical sections were recorded every 0.3 μm. All confocal microscopy images presented in this work are 2D maximum intensity projections of z-stack images (ImageJ 1.47v National Institute of Health,USA).

Personal feedback: A green laser with the appropriate emission filter (530nm) has been used to detect the monomer of the JC1 dye, following FCCP treatment the mitochondrial membrane potential of the cells was eliminated, as demonstrated by the increase of the monomer emission.



JC1 used to assay *in vitro* metabolic effect on hepatocyte and hepatoma cells

Image courtesy of Koukourakis MI et al. Sci Rep. 2016; 6: 30986doi: 10.1038/srep30986. Reproduced under the Creative Commons License <http://creativecommons.org/licenses/by/4.0/>

Koukourakis MI et al. (2016) used the MMP assay kit to stain with JC-1 in NCTC hepatocytes exposed to amifostine (100 µg/ml) over a time course of 20 minutes *in vitro*. Confocal microscopy used to assess mitochondrial membrane potential in the cells.

Serial images confirmed a rapid drop of both green and red fluorescence, one minute after exposure, an effect that was restored to normal at 20 minutes, after a small period of a rebound increase.

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